

V ANCHOR QEA

In Situ Control of Methylmercury Production in Sediments by Redox-Buffering Mineral Amendments

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Hg Biogeochemical Cycle





Biogeochemical Redox Ladder and Hg Methylation

- Ability of microorganisms to methylate Hg linked to possession of *hgc*AB gene cluster
- Present in sulfate-reducing bacteria, as well as some ironreducing and methanogenic bacteria
- To date *hgc*AB gene <u>not</u> found in any manganese-reducers, denitrifiers or aerobic bacteria

[Gilmour et al (2013) Mercury methylation by novel microorganisms from new environments. Environ Sci Technol. 47(20): 11810-20].



Mercury Methylation



Can Sediment Redox Buffering by Manganese Oxides Suppress MeHg Production?





Sediment-Water Slurry Tests

- Investigate effect of MnO₂ on MeHg production in sediment following addition of:
 - labile DOC (acetate, 40 mg/L C)
 - available Hg (HgCl₂, 25 mg/kg)
- Triplicate sediment (150g) + water (500 mL) slurries:
 - control (sediment only)
 - amended (+MnO₂)
- Incubated under N₂ atmosphere in the dark, sampled for THg, MeHg, solution chemistry
- Microbial activity/structure examined by respirometry and qPCR

Sediment

| Analyte | Result (n=3) | SD |
|-----------------------|--------------|-------|
| Total Mercury (mg/kg) | 94.8 | 8.9 |
| TOC (%) | 8.52 | 0.08 |
| lron (mg/kg) | 36,600 | 1,800 |
| Manganese (mg/kg) | 12,700 | 924 |
| Sulfide (mg/kg) | 1.7 | 0.07 |
| Total Solids (%) | 30.0 | 0.2 |

Porewater

| | Analyte | Result |
|------|-------------------|--------|
| | FTHg (ng/L) | 22,900 |
| | FMeHg (ng/L) | 5.86 |
| | Calcium (mg/L) | 106 |
| | Magnesium (mg/L) | 171 |
| | Potassium (mg/L) | 57.7 |
| | Sodium (mg/L) | 1,420 |
| -303 | Chloride (mg/L) | 2,870 |
| 100 | Sulfate (mg/L) | 355 |
| | Alkalinity (mg/L) | 126 |
| | Nitrate-N (mg/L) | 1.69 |
| | Manganese (mg/L) | 0.017 |
| | Iron (mg/L) | < 0.02 |
| | Sulfide (mg/L) | 0.04 |
| | DOC (mg/L) | 6.2 |

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Slurry Test Results

- MeHg in MnO₂-amended microcosms lower than controls by 92 to >99%
- MnO₂ reduced net methylation (%MeHg/THg) by 1-2 orders of magnitude relative to controls
- Redox poising by MnO₂ suppresses MeHg production under conditions otherwise favorable for methylation (sulfate, labile organic matter, available Hg)





Respirometry

- Sediment ± MnO₂ (pyrolusite or birnessite) microcosms inoculated with culture medium (peptone-yeastacetate), incubated under N₂ in the dark
- Continuous monitoring of CO₂ concentration in chamber headspace
- All microcosms showed significant CO₂ production due to microbial activity
- Higher CO₂ production in MnO₂amended microcosms
- MnO₂ amendment of sediment stimulated microbial activity while reducing MeHg production







Microbial Community Census

- PCR and DNA sequencing -phylogenetic composition and major genera present
- Treated and untreated microcosms had similar total eubacterial counts and community composition at the Class level
- Mix of soil and marine bacteria ranging from aerobic to methanogenic
- Dominated by fermentative and anaerobic bacteria
- Iron and manganese reducers present in both control and amended sediment

Phylogenetic Class Distribution



1.0E+06

Sediment Control



Sediment + Pyrolusite

Sediment + Birnessite

Aquarium Mesocosms

- Are MnO₂ amendments effective at suppressing MeHg under more realistic application conditions?
- How long are amendments likely to be effective?
- Aquarium mesocosm tests:
 - Aerated water column
 - 20 cm sediment column
 - MnO₂ added directly to upper sediment layer or applied on top of sediment in a thin layer reactive cover
 - Controls: sediment only and thin layer sand without amendments



Aquarium Microcosm Setup

- Two MnO₂ amendments tested:
 - Pyrolusite (granular, mined)
 - Birnessite (powder, synthesized)
- Two configurations:
 - Direct addition to upper 5 cm of sediment (5 wt%)
 - Mixed in thin layer sand (5 cm, 5 wt%)
- Total of six mesocosms
- Measurements:
 - Overlying water monitored for pH, ORP, SC, Fe, Mn, SO₄, H₂S
 - Porewater and overlying water sampled for MeHg and THg (in replicate)
 - Sediment redox profiles by voltammetry
 - MnO₂ transformation over time (XANES)





Overlying Water

MnO₂ Mixed w/Sediment

Sediment

Overlying Water MnO₂ + Sand

Sediment



Overlying Water Quality Monitoring

0.5



Direct MnO₂ Addition to Sediment





Thin Layer Cover Over Sediment



Arrow indicates date of sampling of porewater for THg/MeHg and voltammetry profiling Horizontal dashed line indicates concentration in site water

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MeHg in Mesocosm Porewater and Overlying Water

- Control tank shows diffusion-reaction depth profile for MeHg
- Birnessite and pyrolusite treatments show reduced MeHg in amended sediment porewater (0-5 cm) and overlying water; ~90% lower than underlying sediment porewater (10-15 cm)
- Sand had lower porewater MeHg concentrations than control, suggests reduced MeHg production, likely due to depletion of sulfate over time
- Birnessite and pyrolusite amended sand reduced both MeHg and THg in overlying water, mixed results for amended sediment porewater 0-5 cm porewater



Direct MnO₂ Addition to Sediment



Effect of MnO₂ on Net MeHg Production

 Reduction in net methylation (as indicated by %MeHg/THg) measured in 0-5 cm porewater relative to control microcosm at 4 months:



| | Pyrolusite | Birnessite | Sand Only |
|------------------|------------|------------|-----------|
| Direct Addition | 69% | 81% | - |
| Thin Layer Cover | 66% | 89% | 65% |





Insights from Redox Profiles (Voltammetry) Direct MnO₂ Addition

- Little penetration of O₂ in sediment (<1 cm)
- Control microcosm showed development of Fe(II) and H₂S (indicators of iron and sulfate reduction) very close to surface (~ 1 cm)
- Both pyrolusite and birnessite inhibited sulfate reduction (no H₂S detected within treatment zone)



Concentration (μ M) or FeS Current (nA)



Insights from Redox Profiles (Voltammetry) Thin Layer Cover Sand

- O₂ penetration to base of sand layer (low O₂ demand)
- Less O₂ penetration in MnO₂-amended sand
- No H₂S detected within sand layer in all 3 microcosms
- Birnessite seems to have a greater depth of influence on redox than pyrolusite



Concentration (μ M) or FeS Current (nA)



Transformation of MnO₂ Amendments with Time

- Mn K-edge XANES used to monitor changes in solid phase manganese speciation over time
- Mn speciation in unamended sediment is predominantly Mn(II), present as rhodochrosite and/or adsorbed species
- In direct addition microcosms, changes in Mn XANES spectra over time indicate partial conversion of original pyrolusite and birnessite to Mn(III) and mixed Mn(II/III) oxides and rhodochrosite





Transformation of MnO₂ Amendments with Time

- In thin layer cover application, Mn XANES spectra show that Mn mineralogy is largely unchanged from original pyrolusite or birnessite over 10 months of microcosm operation
- Contrast with underlying sediment in which Mn is predominantly present as rhodochrosite and/or adsorbed Mn(II)





Transformation of MnO₂ with Time



- MnO₂ converted to Mn(II/III) oxides and rhodochrosite over time
- Redox-buffering Mn phases persist for at least 10 months
- Bulk of added Mn retained in sediment in this system
- Potential for periodic (seasonal and/or tidal) regeneration of MnO₂ by rhodochrosite oxidation in field application



Summary and Conclusions

- Batch slurry experiments document effectiveness of MnO₂ amendments in suppressing net MeHg production in laboratory sediment microcosms
- MnO₂ amendments poise redox and shift predominant microbial activity from sulfate reduction to manganese reduction and suppressing Hg methylation
- In mesocosms, direct addition of MnO₂ amendment to sediment or in thin layer amended sand cover reduced net MeHg production by factor of 3 for pyrolusite and 4-5 for birnessite
- MnO₂ added to sediment was converted to Mn(II/III) oxides and Mn carbonate over time (months) but appears to be retained in solid phase potential for in situ regeneration of MnO₂ through tidal and/or seasonal cycling in field application
- MnO₂ in thin layer sand application converted more slowly than when directly added to sediment – longer effective lifetime

